



Leukotriene B₄ Enzyme Immunoassay Kit

Catalog No. 133-16464

96 Well Kit

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Description

Sapphire's Leukotriene B₄ (LTB₄) EIA kit is a competitive immunoassay for the quantitative determination of LTB₄ in biological samples. This assay is based on the competition between LTB₄ and a LTB₄-alkaline phosphatase tracer for a limited amount of LTB₄-specific antiserum. Because of the competition between LTB₄ in the sample and LTB₄ tracer for the LTB₄ antiserum, the signal obtained with the assay will be inversely proportional to the amount of LTB₄ in each sample. This equilibration is performed in the wells of a 96-well plate which have been coated with mouse monoclonal anti-rabbit IgG, which binds all of the LTB₄ antiserum added to the well. After the incubation step, the plate is washed, and a solution of *para*-nitrophenyl phosphate (*p*NPP), a substrate for alkaline phosphatase, is added. The product of this enzymatic reaction has a distinct yellow color which absorbs at 412 nm, allowing quantification of LTB₄ in each sample.

Introduction

Leukotriene B₄ (LTB₄) is a dihydroxy fatty acid synthesized from arachidonic acid by the combined action of 5-lipoxygenase and LTA₄ hydrolase.¹⁻³ LTB₄ is a potent inflammatory mediator that exerts its effects primarily by binding with low nanomolar affinity to a cell-surface G-protein coupled receptor, BLT₁, on leukocytes.^{4,5} A second low-affinity LTB₄ receptor termed BLT₂ has also been identified but its function is unclear. Binding of LTB₄ to BLT₁ stimulates leukocyte chemotaxis as well as a number of other functions, including aggregation, stimulation of ion fluxes, enhancement of lysosomal enzyme release and superoxide anion production.^{6,7} Plasma levels of LTB₄ increase from <100 pg/ml to >100 ng/ml following leukocyte stimulation.⁸⁻¹⁰ LTB₄ is degraded to inactive metabolites by cytochrome P450 and β-oxidation processes occurring at the ω-end of the molecule.^{7,11}

Precautions

1. Please read all instructions carefully before beginning this assay.
2. Some reagents contain azide, which may react with lead or copper plumbing. If disposing of reagents in the sanitary sewer, please flush with a large volume of water to prevent azide build-up.
3. This kit is for research use only. It is not intended for human or diagnostic use.

Materials Supplied

1. Leukotriene B ₄ Antiserum	1 vial	133-64643
2. Leukotriene B ₄ AP Tracer	1 vial	133-64642
3. Leukotriene B ₄ Standard	1 vial	133-64641
4. Mouse Anti-rabbit IgG Coated Plate	1 plate	133-00008
5. Tris Buffer Concentrate	2 vials	133-00003
6. AP Wash Buffer Concentrate	1 vial	133-00004
7. DEA Buffer Concentrate	1 vial	133-00001
8. <i>p</i> NPP Tablets	5 tablets	133-00002
9. Plate Cover	1 cover	133-00005

Storage

This kit should be stored at -20°C and used before the expiration date printed on the box.

Materials Needed but Not Supplied

1. A source of Millipore water
2. Adjustable pipettors
3. Materials used for sample preparation
4. An orbital shaker
5. A plate reader capable of measuring absorbance between 405-420 nm

Materials needed for sample purification

1. LTB₄ standard to use in determination of recovery
2. 1.0 M HCl, methanol, Millipore water, and ethanol
3. 6 ml SPE C-18 cartridges

Purification Procedure

While the following purification protocol works well for many samples, it is important to be aware that it may not be sufficient for the purification of LTB₄ from all samples.

1. Split samples into two equal parts and place each sample into clean test tubes.
2. Spike one of these sets of samples with LTB₄.
3. Perform the following steps on all samples:
 - a. Acidify the sample to pH <4.0 by the addition of 1.0 M HCl.
 - b. Prepare a C-18 SPE cartridge by conditioning it first with 5 ml of methanol followed by 5 ml Millipore water.
 - c. Apply the sample and allow to flow through the column.
 - d. Rinse the column with 5 ml Millipore water, followed by 5 ml hexane. Allow the column to become dry following the hexane wash.
 - e. Elute the LTB₄ with 5 ml ethanol. If unable to run the assay at this time, store the samples in ethanol at -80°C.
 - f. Dry the sample under a stream of nitrogen. Reconstitute the sample in Tris Buffer and assay both the unspiked and spiked sample in the EIA.

Reagent Preparation

All diluted buffers should be stored at 4°C. When stored in this manner, they will be stable for approximately two months.

1. Tris Buffer

Dilute the contents of one vial of Tris Buffer Concentrate with 90 ml of Millipore water. It is common for the concentrated buffer to contain crystalline salts after thawing. It is important to rinse the vial to obtain any salts that may have precipitated.

2. Wash Buffer

Dilute the 5 ml vial of Wash Buffer to a final volume of 750 ml with Millipore water.

3. DEA Buffer

Dilute the 2.5 ml vial of DEA Buffer Concentrate to a final volume of 25 ml with Millipore water.

4. Leukotriene B₄ Standard

[Note: If assaying culture medium samples that have not been diluted with Tris Buffer, the culture medium rather than Tris Buffer should be used for dilution of the standard curve.]

Equilibrate a pipet tip in ethanol, by repeatedly filling and expelling. Use the equilibrated pipet tip to transfer 100 µl of the LTB₄ standard into a clean test tube. Dilute with 900 µl Millipore water. The concentration of this standard is 50 ng/ml. Label eight glass test tubes #1 - #8. Pipet 900 µl of Tris Buffer into tube #1, and 600 µl into tubes #2 - #8. Transfer 100 µl of the 50 ng/ml standard into tube #1 and vortex to mix. Transfer 400 µl from tube #1 to tube #2. Vortex to mix. Transfer 400 µl from tube #2 to tube #3. Vortex to mix. Continue this process for standard tubes #4 - #8. These diluted standards should be used within twenty-four hours.

5. Leukotriene B₄ Alkaline Phosphatase Tracer

Reconstitute the LTB₄ Alkaline Phosphatase Tracer with 6 ml of Tris buffer. Vortex to mix.

Store this reconstituted tracer at 4°C and use within four weeks.

6. Leukotriene B₄ Antiserum

Reconstitute the LTB₄ Antiserum with 6 ml of Tris Buffer. Vortex to mix.

Store this reconstituted antiserum at 4°C and use within four weeks.

Assay Procedure

[Note: All reagents should be allowed to warm to room temperature before use.]

1. Tris Buffer

Pipet 150 µl Tris Buffer into non-specific binding (NSB) wells, and 100 µl Tris Buffer into zero standard (B₀) wells. If tissue culture medium was used to dilute the standard curve, substitute 100 µl of this same medium for the Tris Buffer in the NSB and B₀ wells.

2. Leukotriene B₄ Standard

Pipet 100 µl of standards into the appropriate wells.

3. Samples

Pipet 100 µl of samples into the appropriate wells. Each sample should be assayed in duplicate or triplicate.

4. Leukotriene B₄ Alkaline Phosphatase Tracer

Pipet 50 µl of tracer into each well except the blank wells and total activity (TA) wells.

5. Leukotriene B₄ Antiserum

Pipet 50 µl of antiserum into each well except the blank wells, TA wells and NSB wells.

Well	Tris Buffer	Std/Sample	Tracer	Antiserum
Blank				
TA			5 µl (at development)	
NSB	150 µl		50 µl	
B ₀	100 µl		50 µl	50 µl
Std/Sample		100 µl	50 µl	50 µl

Pipetting Summary

6. Incubate the plate

Cover each plate with a plate cover and incubate for two hours at room temperature on an orbital shaker.

7. Prepare the pNPP Solution

Dissolve 5 pNPP tablets in 25 ml DEA buffer (25 ml is sufficient to develop 100 wells). *[Note: Reconstituted pNPP is not stable, so we recommend that you make only the amount that you need at any one time.]*

8. Wash the plate

Empty the wells and rinse five times with wash buffer. After the final wash, firmly tap the inverted plate on a paper towel to remove any recalcitrant drops of buffer.

9. Develop the plate

Add 200 µl pNPP solution to each well including blank and TA wells. Add 5 µl of tracer to the TA wells. Cover the plate and allow to develop in the dark on an orbital shaker. This assay typically develops in approximately 60-90 minutes.

10. Read the plate

Wipe the bottom of the plate with a paper towel to remove any finger prints, smudges or dirt which may interfere with obtaining an accurate reading of absorbance. Remove the plate cover, and read the plate at a wavelength of between 405 and 420 nm.

Data Analysis

Most plate readers come with data reduction software that plots data automatically. Alternatively a spreadsheet program can be used. The data should be plotted as %B/B₀ versus log concentration of standard using either a 4-parameter logistic or a log-logit curve fit.

1. Prepare the Data

The following procedure is recommended to prepare the data prior to graphing (subtract the absorbance of the blank wells from all wells on the plate, if not already done).

- a. Average the absorbance readings from the NSB wells (subtract the absorbance of the blank wells from all wells on the plate, if not already done).
- b. Average the absorbance readings from the B₀ wells.
- c. Subtract the average NSB from the average B₀. This is the corrected B₀.

$$\text{Corrected } B_0 = \text{Average } B_0 - \text{Average NSB}$$

- d. Calculate the %B/B₀ for each standard and sample. To do this, subtract the average NSB absorbance from the absorbance and divide by the corrected B₀ (from step c). Multiply by 100 to obtain %B/B₀. Repeat for all wells.

$$\%B/B_0 = \left[\frac{\text{Absorbance} - \text{Average NSB}}{\text{Corrected } B_0} \right] \times 100$$

2. Plot the Standard Curve

Plot %B/B₀ for all standards versus LTB₄ concentration using log (x) and linear (y) axes, and fit the data to a four parameter logistic equation. Alternatively, the data can be linearized using a logit transformation. [*Note: Do not use %B/B₀ in this calculation.*]

$$\text{Logit } (B/B_0) = \ln [B/B_0/(1-B/B_0)]$$

Plot the data as logit(B/B₀) versus log concentration of standard and perform a linear regression fit.

3. Determine the Concentration of your Samples

Calculate the %B/B₀ for each sample. Determine the concentration of each sample using the equations obtained from the analysis of the standard curve. Remember to account for any dilutions made to the sample prior to addition to the well. %B/B₀ values of greater than 80% or less than 20% should be re-assayed as they generally fall outside of the linear range of the standard curve.

4. Correct for Recovery (if purification was performed)

Divide the concentration determined in step #3 by the recovery factor. Correct for any volume changes of the sample which may have occurred during purification.

Recovery Factor =

$$\frac{\text{EIA value of spiked sample (pg/ml)} - \text{EIA value of unspiked sample (pg/ml)}}{\text{Concentration of spike (pg/ml)}}$$

Assay Performance Characteristics

1. Precision

Intra-assay precision was determined by measuring samples containing low, medium, and high concentrations of LTB₄ multiple times in the same assay (eight samples per plate on a total of five plates). Inter-assay precision was determined by measuring low, medium, and high concentrations of the samples eight times using different reagents.

	LTB ₄ (pg/ml)	Intra-assay %CV	Inter-assay %CV
Low	128	8.4	9.8
Medium	320	6.7	6.8
High	800	4.2	5.3

2. Specificity

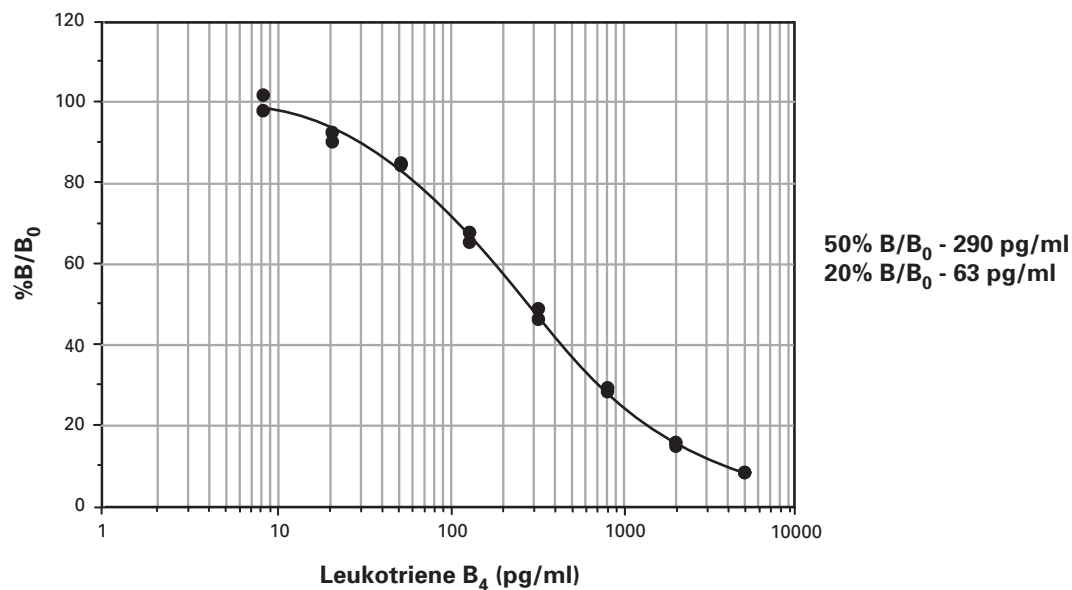
Analyte	Cross-Reactivity	Analyte	Cross-Reactivity
Leukotriene B ₄	100%	20-carboxy Leukotriene B ₄	<0.01%
20-hydroxy Leukotriene B ₄	2.7%	Leukotriene C ₄	<0.01%
5,6-DiHETE	0.07%	Leukotriene D ₄	<0.01%
Glutathione	<0.01%%	19(R)-hydroxy Prostaglandin B ₂	<0.01%

3. Typical Results

The standard curve shown here is an example of data typically produced by this kit. Your results will vary from these, and it is therefore important that you run a standard curve each time you use the kit.

	Raw Data		Average	Corrected
Total Activity	1.247	1.156	1.202	
NSB	0	0	0	
B ₀	0.642	0.669	0.654	0.654
	0645	0.659		

Concentration Std (pg/ml)	Raw Data		Corrected		%B/B ₀	
5000	0.055	0.055	0.055	0.055	8.4	8.4
2000	0.099	0.104	0.099	0.104	15.1	15.9
800	0.186	0.192	0.186	0.192	28.5	29.4
320	0.320	0.303	0.320	0.303	49.0	46.4
128	0.429	0.443	0.429	0.443	65.6	67.8
51.2	0.554	0.557	0.554	0.557	84.5	85.2
20.5	0.605	0.591	0.605	0.591	92.5	90.4
8.2	0.666	0.640	0.666	0.640	101.8	97.9



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13/12/07